

DAFTAR PUSTAKA

- Acland, A., Agarwala, R., Barrett, T., Beck, J., Benson, D. A., Bollin, C., Bolton, E., Bryant, S. H., Canese, K., Church, D. M., Clark, K., DiCuccio, M., Dondoshansky, I., Federhen, S., Feolo, M., Geer, L. Y., Gorelenkov, V., Hoepfner, M., Johnson, M., ... Zbicz, K. (2013). Database resources of the National Center for Biotechnology Information. *Nucleic Acids Research*, *41*(D1). <https://doi.org/10.1093/nar/gks1189>
- Ahrberg, C. D., Ilic, B. R., Manz, A., & Neuzil, P. (2016). Handheld real-time PCR device. *Lab on a Chip*, *16*(3), 586–592. <https://doi.org/10.1039/c5lc01415h>
- Alemu, K. (2014). *Real-Time PCR and Its Application in Plant Disease Diagnostics* (Vol. 27). Online. www.iiste.org
- Amer, H. M., Abd El Wahed, A., Shalaby, M. A., Almajhdi, F. N., Hufert, F. T., & Weidmann, M. (2013). A new approach for diagnosis of bovine coronavirus using a reverse transcription recombinase polymerase amplification assay. *Journal of Virological Methods*, *193*(2), 337–340. <https://doi.org/10.1016/j.jviromet.2013.06.027>
- Anton, D., Burckel, H., Josset, E., & Noel, G. (2015). Three-dimensional cell culture: A breakthrough in vivo. In *International Journal of Molecular Sciences* (Vol. 16, Issue 3, pp. 5517–5527). MDPI AG. <https://doi.org/10.3390/ijms16035517>
- Barbau-Piednoir, E., Botteldoorn, N., Yde, M., Mahillon, J., & Roosens, N. H. (2013). Development and validation of qualitative SYBR®Green real-time PCR for detection and discrimination of *Listeria* spp. and *Listeria monocytogenes*. *Applied Microbiology and Biotechnology*, *97*(9), 4021–4037. <https://doi.org/10.1007/s00253-012-4477-2>
- Birder, L., & Andersson, K.-E. (2013). UROTHELIAL SIGNALING. *Physiol Rev*, *93*, 653–680. <https://doi.org/10.1152/physrev.00030.2012>.-The

- Busk, P. K. (2014). A tool for design of primers for microRNA-specific quantitative RT-qPCR. *BMC Bioinformatics*, *15*(1). <https://doi.org/10.1186/1471-2105-15-29>
- Cai, J., Weiss, M. L., & Rao, M. S. (2004). In search of “stemness.” *Experimental Hematology*, *32*(7), 585–598. <https://doi.org/https://doi.org/10.1016/j.exphem.2004.03.013>
- Chinnadurai, R., Garcia, M. A., Sakurai, Y., Lam, W. A., Kirk, A. D., Galipeau, J., & Copland, I. B. (2014). Actin cytoskeletal disruption following cryopreservation alters the biodistribution of human mesenchymal stromal cells in vivo. *Stem Cell Reports*, *3*(1), 60–72. <https://doi.org/10.1016/j.stemcr.2014.05.003>
- de Kemp, V., de Graaf, P., Fledderus, J. O., Ruud Bosch, J. L. H., & de Kort, L. M. O. (2015). Tissue Engineering for Human Urethral Reconstruction: Systematic Review of Recent Literature. *PLOS ONE*, *10*(2), e0118653-. <https://doi.org/10.1371/journal.pone.0118653>
- Debode, F., Marien, A., Janssen, E., Bragard, C., & Berben, G. (2017). The influence of amplicon length on real-time PCR results. *BASE [En Ligne]*, *21*, 3–11.
- Eling Sasmito, D. K., Kurniawan, R., & Muhimmah, I. (2014). Karakteristik Primer pada Polymerase Chain Reaction (PCR) untuk Sekuensing DNA: Mini Review. In *Seminar Nasional Informatika Medis*.
- Fan, C., Ma, J., Guo, Q., Li, X., Wang, H., & Lu, M. (2013). Selection of Reference Genes for Quantitative Real-Time PCR in Bamboo (*Phyllostachys edulis*). *PLoS ONE*, *8*(2). <https://doi.org/10.1371/journal.pone.0056573>
- Faraldi, M., Mangiavini, L., Conte, C., Banfi, G., Napoli, N., & Lombardi, G. (2022). A novel methodological approach to simultaneously extract high-quality total RNA and proteins from cortical and trabecular bone. *Open Biology*, *12*(5). <https://doi.org/10.1098/rsob.210387>

- Folmes, C. D. L., Kent Arrell, D., Zlatkovic-Lindor, J., Martinez-Fernandez, A., Perez-Terzic, C., Nelson, T. J., & Terzic, A. (2013). Metabolome and metaboproteome remodeling in nuclear reprogramming. *Cell Cycle*, *12*(15), 2355–2365. <https://doi.org/10.4161/cc.25509>
- Garibyan, L., & Avashia, N. (2013). Polymerase chain reaction. *Journal of Investigative Dermatology*, *133*(3), 1–4. <https://doi.org/10.1038/jid.2013.1>
- Gordon Syngai, G., Barman, P., Bharali, R., & Sudip Dey, &. (2013). BLAST: An introductory tool for students to Bioinformatics Applications. *Keanean Journal of Science*, *2*, 67–76.
- Houkpe, B. W., Chenou, F., de Lima, F., & de Paula, E. V. (2021). HRT Atlas v1.0 database: Redefining human and mouse housekeeping genes and candidate reference transcripts by mining massive RNA-seq datasets. *Nucleic Acids Research*, *49*(D1), D947–D955. <https://doi.org/10.1093/nar/gkaa609>
- Hu, L., Wang, J., Zhou, X., Xiong, Z., Zhao, J., Yu, R., Huang, F., Zhang, H., & Chen, L. (2016). Exosomes derived from human adipose mesenchymal stem cells accelerates cutaneous wound healing via optimizing the characteristics of fibroblasts. *Scientific Reports*, *6*. <https://doi.org/10.1038/srep32993>
- Inayatullah, A., Fatmawati, A., & Abdurrahman Munir, M. (2021). *Comparison of Real-Time PCR and Conventional PCR by Identifying Genomic DNA of Bovine and Porcine*.
- Inoue, Y., Kishida, T., Kotani, S. ichiro, Akiyoshi, M., Taga, H., Seki, M., Ukimura, O., & Mazda, O. (2019). Direct conversion of fibroblasts into urothelial cells that may be recruited to regenerating mucosa of injured urinary bladder. *Scientific Reports*, *9*(1). <https://doi.org/10.1038/s41598-019-50388-6>
- Jafari, N. V., & Rohn, J. L. (2022). The urothelium: a multi-faceted barrier against a harsh environment. In *Mucosal Immunology* (Vol. 15, Issue 6, pp. 1127–1142). Springer Nature. <https://doi.org/10.1038/s41385-022-00565-0>
- Jannah, R. M., Naroeni, A., & Novianti, T. (2023). *Bioscaffold from Mouse Embryonic Fibroblasts Maintains the Pluripotency BIOSCAFFOLD FROM*

MOUSE EMBRYONIC FIBROBLAS MAINTAINS THE PLURIPOTENCY OF MOUSE EMBRYONIC STEM CELLS.

- Ka Praja, R., & Rosalina, R. (2021). Perancangan Primer Gen IktB pada *Fusobacterium necrophorum* untuk Analisis PCR (Primer Design of IktB Gene on *Fusobacterium necrophorum* for PCR Assay). *Jurnal Sains Dan Teknologi Peternakan*, 2(2), 47–55. <https://ojs.unsulbar.ac.id/index.php/jstp>
- Kajiwara, M., Aoi, T., Okita, K., Takahashi, R., Inoue, H., Takayama, N., Endo, H., Eto, K., Toguchida, J., Uemoto, S., & Yamanaka, S. (2012). Donor-dependent variations in hepatic differentiation from human-induced pluripotent stem cells. *Proceedings of the National Academy of Sciences of the United States of America*, 109(31), 12538–12543. <https://doi.org/10.1073/pnas.1209979109>
- Kang, M., Kim, H. H., & Han, Y. M. (2014). Generation of bladder urothelium from human pluripotent stem cells under chemically defined serum- and feeder-free system. *International Journal of Molecular Sciences*, 15(5), 7139–7157. <https://doi.org/10.3390/ijms15057139>
- Kim, D., Lee, M., Lee, T. H., Sung, K., Koo, H., & Yoo, K. (2017). Cell culture density affects the stemness gene expression of adipose tissue-derived mesenchymal stem cells. *Biomedical Reports*, 6(3), 300–306. <https://doi.org/10.3892/br.2017.845>
- Kozera, B., & Rapacz, M. (2013). Reference genes in real-time PCR. In *Journal of Applied Genetics* (Vol. 54, Issue 4, pp. 391–406). <https://doi.org/10.1007/s13353-013-0173-x>
- Kralik, P., & Ricchi, M. (2017). A basic guide to real time PCR in microbial diagnostics: Definitions, parameters, and everything. In *Frontiers in Microbiology* (Vol. 8, Issue FEB). Frontiers Research Foundation. <https://doi.org/10.3389/fmicb.2017.00108>
- Kumar, A., & Kaur, J. (2014). Primer Based Approach for PCR Amplification of High GC Content Gene: *Mycobacterium* Gene as a Model . *Molecular Biology International*, 2014, 1–7. <https://doi.org/10.1155/2014/937308>

- Li, Q., Liu, L., Zhang, Q., Liu, S., Ge, D., & You, Z. (2014). Interleukin-17 Indirectly Promotes M2 Macrophage Differentiation through Stimulation of COX-2/PGE2 Pathway in the Cancer Cells. *Crt*, 46(3), 297–306. <https://doi.org/10.4143/crt.2014.46.3.297>
- Lian, H., Roy, E., & Zheng, H. (2016). Protocol for Primary Microglial Culture Preparation. *BIO-PROTOCOL*, 6(21). <https://doi.org/10.21769/bioprotoc.1989>
- Lu, M., & Chai, T. C. (2014). A method to study bladder urothelial cellular function in situ. *Bladder*, 1(1), e3. <https://doi.org/10.14440/bladder.2014.27>
- Marabita, F., De Candia, P., Torri, A., Tegnér, J., Abrignani, S., & Rossi, R. L. (2016). Normalization of circulating microRNA expression data obtained by quantitative real-time RT-PCR. *Briefings in Bioinformatics*, 17(2), 204–212. <https://doi.org/10.1093/bib/bbv056>
- Marsh, T., Pietras, K., & McAllister, S. S. (2013). Fibroblasts as architects of cancer pathogenesis. In *Biochimica et Biophysica Acta - Molecular Basis of Disease* (Vol. 1832, Issue 7, pp. 1070–1078). Elsevier. <https://doi.org/10.1016/j.bbadis.2012.10.013>
- Mohamed Sedek, S. A., Arifin, M. A., & Abdul Munaim, M. S. (2023). qPCR Analysis of Quorum Sensing Genes of *Pseudomonas aeruginosa*. *International Journal of Engineering Technology and Sciences*, 10(1), 43–50. <https://doi.org/10.15282/ijets.10.1.2023.1006>
- Mornkham, T., Wangsomnuk, P. P., Fu, Y. B., Wangsomnuk, P., Jogloy, S., & Patanothai, A. (2013). Extractions of high quality RNA from the seeds of jerusalem artichoke and other plant species with high levels of starch and lipid. *Plants*, 2(2), 302–316. <https://doi.org/10.3390/plants2020302>
- Mubarak, S. M. H., Al-Koofee, D. A. F., Radhi, O. A., Ismael, J. M., & Al-Zubaidi, Z. F. (2020). An optimization and common troubleshooting solving in polymerase chain reaction technique. *Systematic Reviews in Pharmacy*, 11(2), 427–436. <https://doi.org/10.5530/srp.2020.2.63>

- Müller, M., Hermann, P. C., Liebau, S., Weidgang, C., Seufferlein, T., Kleger, A., & Perkhofer, L. (2016). The role of pluripotency factors to drive stemness in gastrointestinal cancer. *Stem Cell Research*, *16*(2), 349–357. <https://doi.org/https://doi.org/10.1016/j.scr.2016.02.005>
- Naroeni, A., Seprianto, S., & Moda, K. F. (2022). In Silico Analysis of SOX2 Gene for Pluripotency Detection at Mouse Embryonic Fibroblas and induced Pluripoten Stem Cell (iPSC). *Nusantara Science and Technology Proceedings*, 1–10. <https://doi.org/10.11594/nstp.2022.2101>
- Narsinh, K. H., Plews, J., & Wu, J. C. (2011a). Comparison of human induced pluripoten and embryonic stem cells: Fraternal or identical twins? In *Molecular Therapy* (Vol. 19, Issue 4, pp. 635–638). <https://doi.org/10.1038/mt.2011.41>
- Narsinh, K. H., Plews, J., & Wu, J. C. (2011b). Comparison of human induced pluripoten and embryonic stem cells: Fraternal or identical twins? In *Molecular Therapy* (Vol. 19, Issue 4, pp. 635–638). <https://doi.org/10.1038/mt.2011.41>
- Onodera, K. (2007). Selection for 3'-End Triplets for Polymerase Chain Reaction Primers. In *Methods in molecular biology* (402nd ed., pp. 61–74). https://doi.org/10.1007/978-1-59745-528-2_3
- Osborn, S. L., Thangappan, R., Luria, A., Lee, J. H., Nolta, J., & Kurzrock, E. A. (2014). Induction of Human Embryonic and Induced Pluripoten Stem Cells Into Urothelium. *Stem Cells Translational Medicine*, *3*(5), 610–619. <https://doi.org/10.5966/sctm.2013-0131>
- Perugini, M., Gallucci, M., & Costantini, G. (2018). A Practical Primer To Power Analysis for Simple Experimental Designs. *International Review of Social Psychology*. <https://doi.org/10.5334/irsp.181>
- Pierce, K. E., Aquiles Sanchez, J., Rice, J. E., & Wangh, L. J. (2005). Linear-After-The-Exponential (LATE)-PCR: Primer design criteria for high yields of specific single-stranded DNA and improved real-time detection. In *PNAS* (Vol. 102). www.pnas.org/cgi/doi/10.1073/pnas.0501946102

- Pinto, J. P., Kalathur, R. K., Oliveira, D. V., Barata, T., Machado, R. S. R., Machado, S., Pacheco-Leyva, I., Duarte, I., & Futschik, M. E. (2015). StemChecker: a web-based tool to discover and explore stemness signatures in gene sets. *Nucleic Acids Research*, 43(W1), W72–W77. <https://doi.org/10.1093/nar/gkv529>
- Punca, S., Karakteristik, :, Dan, P., Budiman, A., Staf, H., Bagian, P., Fakultas, B., Ukrida, K., & Korespondensi, A. (2016). Tinjauan Pustaka. In *J. Kedokt Meditek* (Vol. 22, Issue 60).
- Quan, P. L., Sauzade, M., & Brouzes, E. (2018). DPCR: A technology review. In *Sensors (Switzerland)* (Vol. 18, Issue 4). MDPI AG. <https://doi.org/10.3390/s18041271>
- Raab, S., Klingenstein, M., Liebau, S., & Linta, L. (2014). A Comparative View on Human Somatic Cell Sources for iPSC Generation. In *Stem Cells International* (Vol. 2014). Hindawi Limited. <https://doi.org/10.1155/2014/768391>
- Ragni, E., Viganò, M., Rebullà, P., Giordano, R., & Lazzari, L. (2013). What is beyond a qRT-PCR study on mesenchymal stem cell differentiation properties: How to choose the most reliable housekeeping genes. *Journal of Cellular and Molecular Medicine*, 17(1), 168–180. <https://doi.org/10.1111/j.1582-4934.2012.01660.x>
- Rao, X. , H. X. , Z. Z. , & L. X. (2013). An improvement of the $2^{-\Delta\Delta CT}$ method for quantitative real-time polymerase chain reaction data analysis. *Biostatistics, Bioinformatics and Biomathematics.*, 3(3), 71–85.
- Rasool, A., Qu, Q., Wang, Y., & Jiang, Q. (2022). Bio-Constrained Codes with Neural Network for Density-Based DNA Data Storage. *Mathematics*, 10(5). <https://doi.org/10.3390/math10050845>
- Ray, D. L., & Johnson, J. C. (2014). Validation of reference genes for gene expression analysis in olive (*Olea europaea*) mesocarp tissue by quantitative real-time RT-PCR. *BMC Research Notes*, 7(1). <https://doi.org/10.1186/1756-0500-7-304>

- Redshaw, N., Wilkes, T., Whale, A., Cowen, S., Huggett, J., & Foy, C. A. (2013). A comparison of miRNA isolation and RT-qPCR technologies and their effects on quantification accuracy and repeatability. *BioTechniques*, *54*(3), 155–164. <https://doi.org/10.2144/000114002>
- Ruiz-Villalba, A., van Pelt-Verkuil, E., Gunst, Q. D., Ruijter, J. M., & van den Hoff, M. J. (2017). Amplification of nonspecific products in quantitative polymerase chain reactions (qPCR). *Biomolecular Detection and Quantification*, *14*, 7–18. <https://doi.org/10.1016/j.bdq.2017.10.001>
- Siddiqui, S. (2015). Therapeutic Potential of Totipotent, Pluripoten and Multipotent Stem Cells. *MOJ Cell Science & Report*, *2*(5). <https://doi.org/10.15406/mojcsr.2015.02.00041>
- Singh, R. R. (2020). Next-Generation Sequencing in High-Sensitive Detection of Mutations in Tumors: Challenges, Advances, and Applications. In *Journal of Molecular Diagnostics* (Vol. 22, Issue 8, pp. 994–1007). Elsevier B.V. <https://doi.org/10.1016/j.jmoldx.2020.04.213>
- Suh, T. C., Amanah, A. Y., & Gluck, J. M. (2020). Electrospun scaffolds and induced pluripoten stem cell-derived cardiomyocytes for cardiac tissue engineering applications. In *Bioengineering* (Vol. 7, Issue 3, pp. 1–22). MDPI AG. <https://doi.org/10.3390/bioengineering7030105>
- Suzuki, K., Koyanagi-Aoi, M., Uehara, K., Hinata, N., Fujisawa, M., & Aoi, T. (2019). Directed differentiation of human induced pluripoten stem cells into mature stratified bladder urothelium. *Scientific Reports*, *9*(1). <https://doi.org/10.1038/s41598-019-46848-8>
- Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K., & Yamanaka, S. (2007). Induction of Pluripoten Stem Cells from Adult Human Fibroblasts by Defined Faktors. *Cell*, *131*(5), 861–872. <https://doi.org/https://doi.org/10.1016/j.cell.2007.11.019>
- Teo, A. K. K., Lau, H. H., Valdez, I. A., Dirice, E., Tjora, E., Raeder, H., & Kulkarni, R. N. (2016). Early Developmental Perturbations in a Human Stem

- Cell Model of MODY5/HNF1B Pancreatic Hypoplasia. *Stem Cell Reports*, 6(3), 357–367. <https://doi.org/10.1016/j.stemcr.2016.01.007>
- Thanan, R., Murata, M., Ma, N., Hammam, O., Wishahi, M., El Leithy, T., Hiraku, Y., Oikawa, S., & Kawanishi, S. (2012). Nuclear Localization of COX-2 in relation to the Expression of Stemness Markers in Urinary Bladder Cancer. *Mediators of Inflammation*, 2012, 165879. <https://doi.org/10.1155/2012/165879>
- Thijs, S., De Beeck, M. O., Beckers, B., Truyens, S., Stevens, V., Van Hamme, J. D., Weyens, N., & Vangronsveld, J. (2017). Comparative evaluation of four bacteria-specific primer pairs for 16S rRNA gene surveys. *Frontiers in Microbiology*, 8(MAR). <https://doi.org/10.3389/fmicb.2017.00494>
- Wan, Z., Zhang, Y., He, Z., Liu, J., Lan, K., Hu, Y., & Zhang, C. (2016). A melting curve-based multiplex RT-qPCR assay for simultaneous detection of four human coronaviruses. *International Journal of Molecular Sciences*, 17(11). <https://doi.org/10.3390/ijms17111880>
- Wang, X., Nakamoto, T., Dulińska-Molak, I., Kawazoe, N., & Chen, G. (2016). Regulating the stemness of mesenchymal stem cells by tuning micropattern features. *J. Mater. Chem. B*, 4(1), 37–45. <https://doi.org/10.1039/C5TB02215K>
- Wu, X. R., Kong, X. P., Pellicer, A., Kreibich, G., & Sun, T. T. (2009). Uroplakins in urothelial biology, function, and disease. In *Kidney International* (Vol. 75, Issue 11, pp. 1153–1165). <https://doi.org/10.1038/ki.2009.73>
- Zhang, D., Sun, F., Yao, H., Wang, D., Bao, X., Wang, J., & Wu, J. (2022). Generation of Urothelial Cells from Mouse-Induced Pluripoten Stem Cells. *International Journal of Stem Cells*, 15(4), 347–358. <https://doi.org/10.15283/ijsc21250>
- Zhang, S. (2014). SOX2, a key faktor in the regulation of pluripotency and neural differentiation. *World Journal of Stem Cells*, 6(3), 305. <https://doi.org/10.4252/wjsc.v6.i3.305>

Zhao, F., Maren, N. A., Kosentka, P. Z., Liao, Y. Y., Lu, H., Duduit, J. R., Huang, D., Ashrafi, H., Zhao, T., Huerta, A. I., Ranney, T. G., & Liu, W. (2021). An optimized protocol for stepwise optimization of real-time RT-PCR analysis. *Horticulture Research*, 8(1). <https://doi.org/10.1038/s41438-021-00616-w>